

**ab138873**

**Acetylcholinesterase  
Assay Kit (Fluorometric -  
Red)**

**Instructions for Use**

For the detection of Acetylcholinesterase activity in blood, cell extracts, and in other solutions.

This product is for research use only and is not intended for diagnostic use.



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# 1. Introduction

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Acetylcholinesterase (AChE) is one of the most crucial enzymes for nerve response and function. AChE degrades the neurotransmitter acetylcholine (ACh) into choline and acetic acid. It is mainly found at neuromuscular junctions and cholinergic synapses in the central nervous system, where its activity serves to terminate the synaptic transmission. AChE inhibitors are among the key drugs approved for Alzheimer's disease (AD) and myasthenia gravis.

ab138873 Acetylcholinesterase Assay Kit (Fluorometric -Red) provides one of the most sensitive methods for detecting AChE activity or screening AChE inhibitors in red fluorescence window. The kit uses AbRed Indicator to quantify the choline produced from the hydrolysis of acetylcholine by AChE through choline oxidase-mediated enzyme coupling reactions. It can be used for monitoring and quantifying the AChE activity in blood, cell extracts or other solutions. The fluorescence intensity of AbRed Indicator is used to measure the amount of choline formed, which is proportional to the AChE activity. The kit is an optimized "mix and read" assay that provides a simple one-step fluorometric assay to detect as little as 0.01 mU AChE in a 100  $\mu$ l assay volume (0.1 mU/ml). Its signal can be easily read with a fluorescence microplate reader at Ex/Em = ~540/590 nm or an absorbance microplate reader at ~575 nm.

## Kit Key Features

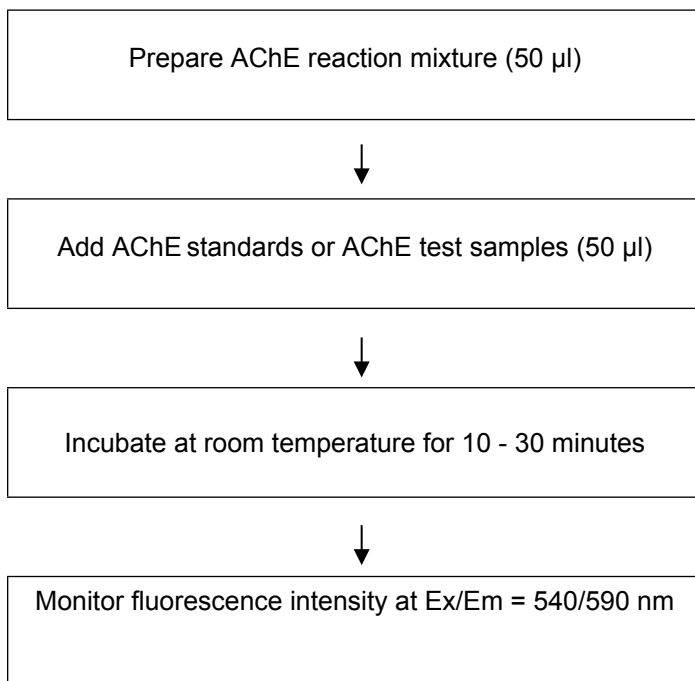
- **Broad Application:** Can be used to quantify acetylcholinesterase in solutions and in cell extracts.
- **Sensitive:** Detect as low as 0.01 mU of acetylcholinesterase in solution.
- **Continuous:** Easily adapted to automation without a separation step.
- **Convenient:** Formulated to have minimal hands-on time.

This product does not differentiate between acetylcholinesterase (AChE) or butyrylcholinesterase (BChE) activity as both enzymes can hydrolyze acetylcholine.

## 2. Protocol Summary

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*Summary for One 96-well Plate*



*Note: Thaw all the kit components to room temperature before starting the experiment.*

### 3. Kit Contents

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<b>Components</b>	<b>Amount</b>
Component A: AbRed Indicator	1 vial
Component B: Acetylcholinesterase Probe	2 bottles (lyophilized powder)
Component C: Acetylthiocholine	1 vial
Component D: Acetylcholinesterase Standard	1 vial (5 units)
Component E: Assay Buffer	1 bottle (10 ml)
Component F: Dilution Buffer	1 bottle (10 ml)
Component G: DMSO	1 vial (100 $\mu$ L)

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## 4. Storage and Handling

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Keep at -20°C. Avoid exposure to light.

## 5. Additional Materials Required

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- 96 or 384-well solid black microplates
- Fluorescence microplate reader
- Optional: AchE specific inhibitor. We recommend:
  - Territrem B (ab144370)
  - Donepezil hydrochloride (ab120763)
  - Cyclopenin (ab144233)



## 6. Assay Protocol

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**Note:** *This protocol is for one 96 - well plate.*

### A. Prepare Stock Solutions

1. 250X AbRed Indicator stock solution: Add 40  $\mu\text{L}$  of DMSO (Component G) into the vial of AbRed Indicator (Component A) to make 250X AbRed Indicator stock solution.

*Note: The unused AbRed Indicator stock solution should be divided into single use aliquots. Store at  $-20\text{ }^{\circ}\text{C}$  and keep from light.*

2. Acetylcholinesterase standard stock solution: Add 100  $\mu\text{L}$  of Assay Buffer (Component E) into the vial of acetylcholinesterase standard (Component D) to make a 50 units/ml acetylcholinesterase standard stock solution

*Note: The unused acetylcholinesterase standard stock solution should be divided into single use aliquots and stored at  $-20\text{ }^{\circ}\text{C}$ .*

3. Acetylcholine stock solution: Add 100  $\mu\text{L}$  of Assay Buffer (Component E) into the vial of Acetylcholine

(Component C) to make 1000X acetylcholine stock solution.

*Note: The unused acetylcholine stock solution should be divided into single use aliquots and stored at -20 °C.*

## **B. Prepare acetylcholinesterase assay mixture**

1. Add 5 ml of Assay Buffer (Component E) to the bottle of Acetylcholinesterase Probe (Component B) and mix well.
2. Add 5  $\mu$ L of 1000X acetylcholine stock solution into the bottle of Acetylcholinesterase Probe mixture and mix well.
3. Add 20  $\mu$ l of 250X AbRed Indicator stock solution into the bottle of Acetylcholinesterase Probe mixture to make the acetylcholinesterase assay mixture before running the assay.

*Note: The acetylcholinesterase assay mixture should be used promptly and kept from light. The assay background would increase with longer storage time.*

**C. Prepare serially diluted acetylcholinesterase standards (0 to 100 mU/ml):**

1. Add 20  $\mu$ l of 50 units/ml acetylcholinesterase standard stock solution to 980  $\mu$ l Dilution Buffer (Component F) to generate 1000 mU/ml acetylcholinesterase standard solution.

*Note: Diluted acetylcholinesterase standard solution is unstable and should be used within 4 hours.*

2. Take 200  $\mu$ l of 1000 mU/ml acetylcholinesterase standard solution to perform 1:10 and 1:3 serial dilutions to get 100, 30, 10, 3, 1, 0.3, 0.1 and 0 mU/ml serially diluted acetylcholinesterase standards.

3. Add serially diluted acetylcholinesterase standards and/or acetylcholinesterase containing test samples into a solid black 96-well microplate as described in Tables 1 and 2.

*Note: Treat cells or tissue samples as desired.*

BL	BL	TS	TS	....	....						
AS1	AS1	....	....	....	....						

AS2	AS2										
AS3	AS3										
AS4	AS4										
AS5	AS5										
AS6	AS6										
AS7	AS7										

**Table 1.** Layout of acetylcholinesterase standards and test samples in a solid black 96-well microplate.

*Note: AS= Acetylcholinesterase Standards; BL=Blank Control; TS=Test Samples*

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<b>Acetylcholinesterase Standards</b>	<b>Blank Control</b>	<b>Test Sample</b>
Serial Dilutions*: 50 µl	Assay Buffer: 50 µl	50 µl

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**Table 2.** Reagent composition for each well.

*\*Note: Add the serial dilutions of acetylcholinesterase standard from 1 to 1000 mU/ml into wells from AS1 to AS7 in duplicate.*

**D. Run acetylcholinesterase assay:**

1. Add 50  $\mu$ l of acetylcholinesterase assay mixture into each well of acetylcholinesterase standard, blank control, and test samples to make the total acetylcholinesterase assay volume of 100  $\mu$ l/well.

*Note: For a 384-well plate, add 25  $\mu$ l of sample and 25  $\mu$ l of acetylthiocholine reaction mixture in each well.*

2. Incubate the reaction for 10 to 30 minutes at room temperature, protected from light.
3. Monitor the fluorescence increase with a fluorescence microplate reader at Ex/Em = 540/590 nm.

NOTE: Butyrylcholinesterase (BChE) present in the sample can convert acetylcholine and lead to false positives. We recommend using a specific acetylcholinesterase as a control:

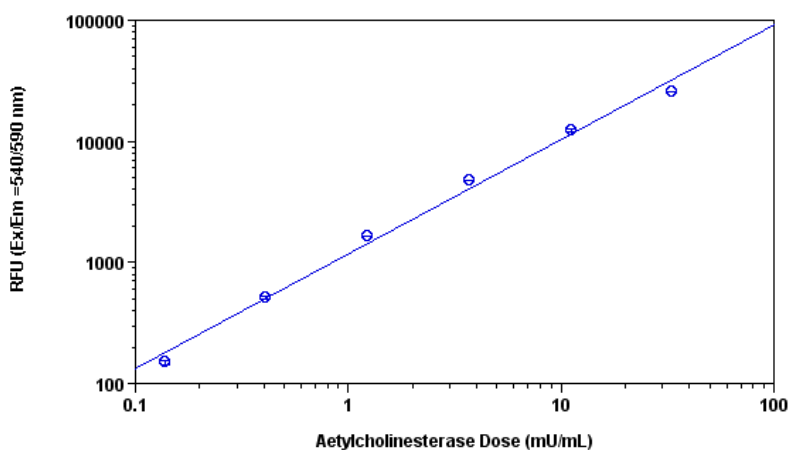
- Territrem B (ab144370)
- Donepezil hydrochloride (ab120763)
- Cyclopenin (ab144233)

## 7. Data Analysis

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The fluorescence in blank wells (with the Dilution Buffer only) is used as a control, and is subtracted from the values for those wells with the acetylcholinesterase reactions. An acetylcholinesterase standard curve is shown in Figure 1.

*Note: The fluorescence background increases with time, thus it is important to subtract the fluorescence intensity value of the blank wells for each data point.*



**Figure 1.** Acetylcholinesterase dose response was measured in a solid black 96-well plate with ab138873 using a fluorescence microplate reader. As low as 0.01 mU/well (0.1 mU/ml) acetylcholinesterase can be detected with 20 minutes incubation (n=3).

## 8. Troubleshooting

<b>Problem</b>	<b>Reason</b>	<b>Solution</b>
Assay not working	Assay buffer at wrong temperature	Assay buffer must not be chilled - needs to be at RT
	Protocol step missed	Re-read and follow the protocol exactly
	Plate read at incorrect wavelength	Ensure you are using appropriate reader and filter settings (refer to datasheet)
	Unsuitable microtiter plate for assay	Fluorescence: Black plates (clear bottoms); Luminescence: White plates; Colorimetry: Clear plates. If critical, datasheet will indicate whether to use flat- or U-shaped wells
Unexpected results	Measured at wrong wavelength	Use appropriate reader and filter settings described in datasheet
	Samples contain impeding substances	Troubleshoot and also consider deproteinizing samples
	Unsuitable sample type	Use recommended samples types as listed on the datasheet
	Sample readings are outside linear range	Concentrate/ dilute samples to be in linear range



<b>Problem</b>	<b>Reason</b>	<b>Solution</b>
Samples with inconsistent readings	Unsuitable sample type	Refer to datasheet for details about incompatible samples
	Samples prepared in the wrong buffer	Use the assay buffer provided (or refer to datasheet for instructions)
	Samples not deproteinized (if indicated on datasheet)	Use the <b>10kDa spin column (ab93349)</b> or <b>Deproteinizing sample preparation kit (ab93299)</b>
	Cell/ tissue samples not sufficiently homogenized	Increase sonication time/ number of strokes with the Dounce homogenizer
	Too many freeze-thaw cycles	Aliquot samples to reduce the number of freeze-thaw cycles
	Samples contain impeding substances	Troubleshoot and also consider deproteinizing samples
	Samples are too old or incorrectly stored	Use freshly made samples and store at recommended temperature until use
Lower/ Higher readings in samples and standards	Not fully thawed kit components	Wait for components to thaw completely and gently mix prior use
	Out-of-date kit or incorrectly stored reagents	Always check expiry date and store kit components as recommended on the datasheet
	Reagents sitting for extended periods on ice	Try to prepare a fresh reaction mix prior to each use
	Incorrect incubation time/ temperature	Refer to datasheet for recommended incubation time and/ or temperature
	Incorrect amounts used	Check pipette is calibrated correctly (always use smallest volume pipette that can pipette entire volume)

Standard curve is not linear	Not fully thawed kit components	Wait for components to thaw completely and gently mix prior use
	Pipetting errors when setting up the standard curve	Try not to pipette too small volumes
	Incorrect pipetting when preparing the reaction mix	Always prepare a master mix
	Air bubbles in wells	Air bubbles will interfere with readings; try to avoid producing air bubbles and always remove bubbles prior to reading plates
	Concentration of standard stock incorrect	Recheck datasheet for recommended concentrations of standard stocks
	Errors in standard curve calculations	Refer to datasheet and re-check the calculations
	Use of other reagents than those provided with the kit	Use fresh components from the same kit

**For further technical questions please do not hesitate to contact us by email ([technical@abcam.com](mailto:technical@abcam.com)) or phone (select “contact us” on [www.abcam.com](http://www.abcam.com) for the phone number for your region).**



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